# **Expert Opinion**

- Introduction
- The physiological role of TF
- The extrinsic coagulation cascade in thrombosis
- Non-thrombotic effect of TF-thrombin pathway
- Anticoagulation therapies
- Conclusion
- **Expert opinion**

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## The multivalent activity of the tissue factor-thrombin pathway in thrombotic and non-thrombotic disorders as a target for therapeutic intervention

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Importance of the field: Tissue factor (TF) is the key initiator of the coagulation cascade. The exposure of subendothelial TF after vessel injury to blood is a critical step in hemostasis and in the pathogenesis of arterial and venous thrombotic disorders. Moreover, an additional role for TF overexpression and subsequent generation of TF:FVIIa complex, FXa and thrombin have been recently emerged, contributing in non-thrombotic manifestations such as inflammation, cancer growth and fibrosis.

Areas covered in this review: The multivalent role of TF and the above mentioned proteases in disease is reviewed, with focus on their implication in non-thrombotic disorders, as suggested by clinical and experimental data. Moreover, potential therapeutic interventions using anticoagulation agents are discussed.

What the reader will gain: A better understanding of the pathogenic role of the TF-thrombin pathway in the pathogenesis of disease and the effect of anticoagulants in the treatment of such disorders.

Take home message: The TF-thrombin pathway, apart from the initiation of hemostasis and thrombosis, exert intracellular signaling activity through protease-activated receptors, participating in inflammation and tumor biology. Both low-molecular-weight heparins and recently developed anticoagulants rise as candidates for the modification of biological functions associated with disorders like sepsis, ischemia-reperfusion or cancer growth and metastasis.

Keywords: cancer, inflammation, TF, thrombin, thrombosis

Expert Opin. Ther. Targets [Early Online]

#### 1. Introduction

In 1905 Morawitz stated a clotting theory, proposing a crucial and calciumdependent role for thrombokinase in the conversion of prothrombin (thrombogen, as he named it) to thrombin. Later, in 1908, the term tissue thromboplastin was used instead of thrombokinase by Nolf, and even later, in 1935, the term tissue factor (TF) was proposed by Howell. Although various theories regarding clotting had been proposed, the clotting theory of Morawitz influenced the research in the field for many decades. In 1977, it was shown that the complex TF-factor VII (FVII) activated both factor X (FX) and IX (FIX), holding a significant role in clot formation. Thus, 90 years later, in 1995, Rapaport and Rao pointed to TF as the main factor able to trigger coagulation in vivo. They wrote: "..the binding of factor VII



#### Article highlights.

- The increased expression of TF by leukocytes, endothelium or cancer cells initiates the activation of the coagulation cascade that leads to thrombin generation.
- The TF-thrombin pathway is implicated in vascular microangiopathy and in venous and arterial thrombosis.
- The proteases of the extrinsic coagulation system exert signaling through PARs, regulating several inflammatory responses and tumor growth and metastasis.
- Experimental and clinical data provide evidence for the effectiveness of anticoagulation-based strategies in the treatment of inflammatory and malignant disorders.
- Several recently developed agents with favorable safety profile and tolerability constitute promising alternative treatment strategies for the treatment of both thrombotic and non-thrombotic disorders.

This box summarizes key points contained in the article

to TF and the subsequent reactions so triggered play a 'prima ballerina' role in the initiation of coagulation" [1,2].

During the period 1980 - 2000, the exons/introns organization of the TF gene and the characterization of the structure of TF protein contributed to the resolution of various questions regarding the expression of TF by different cells and tissues, its natural inhibitor, TF pathway inhibitor (TFPI), [2], the function of TF isoforms, the characterization of bloodborne TF and the appropriate or not induction of TF expression under several clinical conditions.

Despite the characterization of TF structure and function, no human disorder that depends on a genetic defect of this factor has been described. However, it is currently established that the activation of TF-FVII complex has, apart from its procoagulant activity, a signaling activity too. Proteaseactivated receptors (PARs) [3] mediate the regulation of nonthrombotic functions such as inflammation, angiogenesis, tumor growth and fibrosis by the proteases of this pathway, namely TF-FVIIa complex, FXa and thrombin.

Since the TF pathway constitutes a pleiotropic system involved in various disorders, thrombotic or non-thrombotic, it is reasonable for it to be currently considered as a multivalent target for future therapeutic intervention. The field of selective targeting of either TF-VII signaling or TF pathway inducers is rapidly evolving and novel and safe agents are available in experimental trials and clinical use.

#### 2. The physiological role of TF

#### 2.1 TF structure

TF is a 47 kDa transmembrane glycoprotein that demonstrates high homology in secondary and tertiary structure with interferon  $\gamma$  receptors and is a member of the human class II cytokine receptor family [4]. TF is encoded by a 12.4 kb gene on chromosome 1, which is organized into six exons [5]. The main TF mRNA transcript is comprised by 2.3 kb [5], while the existence of larger transcripts [6,7] with

no physiological relevance has been reported [8]. Recently, a functional alternative spliced variant of TF (asTF) has been identified by Bogdanov and colleagues [9]; asTF transcript lacks exon 5 which encodes for the transmembrane domain, resulting in a soluble shorter protein of 206 amino acids [9].

#### 2.2 TF expression

The plethora of activating binding sites of TF promoter region indicates multipotency in gene expression, permitting TF expression in a variety of cells under different stimuli and conditions (Figure 1). In addition, methylation of the promoter could prevent expression in certain cells such as T and B lymphocytes. More specifically, TF is expressed in cells of the vascular wall, astrocytes, cells lining organs, fibroblasts, smooth muscle cells and adipocytes [10,11]. TF is not expressed in endothelial cells under physiological conditions [12], preventing the unintended and potentially dangerous activation of blood coagulation. However, various subendothelial cells express significant amounts of TF, able to activate the extrinsic coagulation cascade after the disruption of vessel wall integrity. Inflammatory molecules (TNF-α, IL-1β, IL-8, C5a) [13-17] and mitogens (VEGF) [18] are able to induce TF expression in endothelial cells, monocytes and peripheral polymorphonuclear leukocytes (Figure 1). The promoter region of TF possesses two binding sites for the transcription factor activator protein-1 (AP-1), one nuclear factor KB site, three early growth response gene-1 (Egr-1) and five specific protein-1 (Sp1) sites [19-21]. The presence of a CpG island on the first part of the TF promoter sequence provides a methylating site [5].

#### 2.3 Blood-borne TF

Until recently, unstimulated peripheral blood cells were considered unable to express TF, supporting the theory of the haemostatic envelope of blood vessels. However, emerging evidence indicates the presence of circulating TF in blood (also called blood-borne TF). Blood-borne TF originates from three possible sources: peripheral blood cells [10,13,14,16,17], circulating microparticles (MP) containing TF [22] and the soluble alternative spliced variant of TF [8]. Peripheral blood monocytes and neutrophils can express TF when stimulated by different factors such as LPS, TNF-α, and C5a [13,14,16,17]. Furthermore, small lipoprotein structures formed by vesiculation of cell membrane of endothelial and peripheral blood cells (microparticles (MP)) were found to contain TF [22]. Additionally, asTF constitutes a significant part of bloodborne TF [9]. Despite the fact that its procoagulant activity is not clearly defined [9,23], as TF seems to be a functional variant that can activate FVII for further signalling while its contribution in angiogenesis has been recently described [24].

#### 2.4 Haemostatic activity of TF

TF is released in blood from tissues in the event of vessel wall injury. Furthermore, activated endothelial cells and peripheral blood cells express TF after stimulation through several



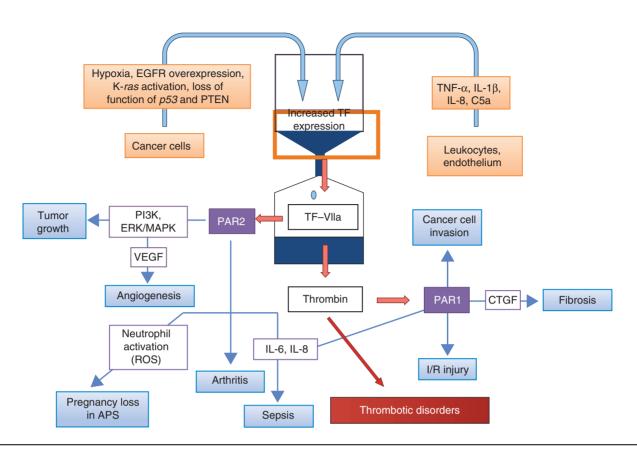


Figure 1. TF in the bottleneck of disease pathogenesis. Several stimuli induce the expression of TF in endothelial cells and/ or blood cells (leukocytes, platelets). Moreover, cancer cells express high levels of TF, in response to hypoxia or due dysregulation of oncogenes and/or tumor suppressor genes. The increased levels of TF result in the activation of the extrinsic coagulation cascade, leading to venous thrombosis. In addition, the proteases of this cascade activate PARs, participating in the pathogenesis of inflammation, tumor growth, angiogenesis and fibrosis.

APS: Antiphospholipid syndrome; CTGF: Connective tissue growth factor; ERK: Extracellular-signal-regulated kinase; I/R: Ischemia-reperfusion; PAR: Protease activated receptor; PTEN: Phosphatase and tensin homologue; ROS: Reactive oxygen species.

mediators. TF acts as the main in vivo initiator of coagulation, when it comes into contact with blood [25]. The activation of the extrinsic coagulation system in a sequential cascade of events plays a critical role in hemostasis. The extrinsic coagulation cascade is initiated when TF, localized on cell surface, binds and activates FVII. The TF-FVIIa complex further activates both vitamin-K-dependent zymogens, FIX and FX. FXa in turn, and in conjunction with FVa, converts prothrombin to thrombin [26]. The generated thrombin, even though not sufficient for the formation of fibrin, amplifies the initial thrombin signal. Thrombin binds to platelets that adhere at the site of tissue injury and expose on their membranes phosphatidylserine, an anionic phospholipid involved in the further activation of the coagulation cascade. The recruitment of platelets enables the assembly of the intrinsic tenase (FIXa/FVIIIa) and prothrombinase (FXa/FVa) complexes, further activating FX and resulting in the conversion of prothrombin into thrombin, respectively [27]. FVII, FX and thrombin belong to the family of serine proteases and signal through proteolytic cleavage/activation of other

proteins/receptors, participating in several functions other than thrombosis (Figure 2) [28]. The coagulation cascade is regulated by natural anticoagulants such as protein C, protein S and TFPI.

#### 3. The extrinsic coagulation cascade in thrombosis

Inappropriate TF expression is implicated in the pathogenesis of arterial and venous thrombotic disorders. It is suggested that the exposure of subendothelial TF to blood and/or the induction of TF expression in endothelial and blood cells is the critical step for the pathologic activation of coagulation that characterizes arterial and venous thrombosis, respectively (Figures 1 and 2) [26].

#### 3.1 Atherothrombosis

Cardiovascular disorders constitute the leading cause of mortality in the developed world, having a major effect on public health. The rupture of an atherosclerotic plaque and the

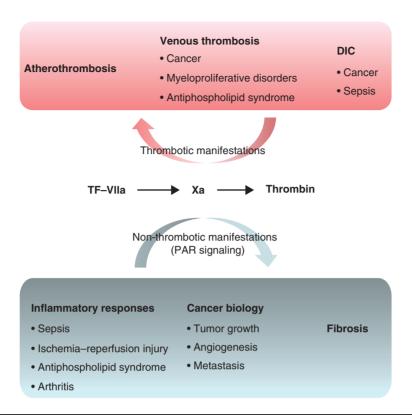


Figure 2. The TF-thrombin pathway is implicated in thrombotic and non-thrombotic disorders. The activation of the extrinsic coagulation cascade and the generation of thrombin is the key event in the pathogenesis of venous thrombosis and disseminated intravascular coagulation (DIC). This cascade is further implicated in arterial thrombosis by stabilizing the initially formed platelet-rich thrombi. Moreover, TF-FVIIa complex, FXa and thrombin signaling through protease activated receptors (PARs) is involved in the pathogenesis of inflammatory disorders, including sepsis, ischemia-reperfusion injury, antiphospholipid syndrome and arthritis, cancer (participating in metastasis, tumor growth and angiogenesis), and probably in fibrotic disorders.

formation of platelet rich thrombi represent the initial event in atherothrombosis. The subsequent release of TF, which is highly expressed in the subendothelial tissue [12,29] activates blood coagulation and results in fibrin deposition on the platelet surface and stabilization of thrombus [30]. Whether blood-born TF, localized on blood cells or MP, contributes in this process is still a matter of debate [20]. Even though the levels of circulating TF have been associated with acute myocardial infarction [31], data extracted from ex vivo studies or in vivo animal models of arterial thrombosis remain controversial. TF derived from the vessel wall and not from blood cells was involved in thrombosis in healthy mice after carotid artery injury [32], while a laser-induced injury model of mouse cremaster arterioles implicated circulating TF in thrombosis [33].

#### 3.2 Venous thrombosis

Available evidence points towards aberrant TF expression among the main pathogenic mechanisms for the majority of deep vein thrombosis (DVT) and pulmonary embolism (PE) events, while the inherited disorders of coagulation and fibrinolytic systems are currently considered as predisposing

factors. Circulating TF, expressed on MP derived from activated or apoptotic cells or blood cells, is thought to play the key role in venous thrombosis, since vessel-wall damage is not observed in thrombotic events in this compartment of systemic circulation [26]. Concerning procoagulant MP, the identification of increased levels of TF-positive MP in patients with cancer [34-37] or myeloproliferative disorders (MPD) [38] suggests their implication in thrombosis.

#### 3.2.1 Cancer and thrombosis

Thromboembolic disease is a common complication in patients with cancer. It constitutes a leading cause of death in patients with adenocarcinoma, while about 20% of the total burden of venous thromboembolism is associated with malignancies [39]. Several experimental studies tried to elucidate the pathogenic mechanism of thrombosis in cancer since the original description of the association between cancer and coagulation by Armand Trousseau.

Increased TF expression in both stromal and neoplastic cells [40] has been demonstrated in many malignancies, including colorectal [41], pancreatic [42], prostate [37] and breast cancer [43], and is involved in the prothrombotic



state of patients with malignancies [39]. Enhanced TF expression by cancer cells depends on hypoxia or genetic transformation, including overexpression of the oncogenic EGFR, activation of K-ras oncogene or loss of function of the tumor suppressors p53 and phosphatase and tensin homologue (PTEN) [40].

Increased levels of TF-bearing MP have been detected in patients suffering from malignancy, especially pancreatic and breast cancer [34-37]. Interestingly, circulating TF has been proposed as a biomarker for the identification of patients at increased risk for thromboembolism [44]. Moreover, a correlation between TF-bearing MP levels and overt disseminated intravascular coagulation (DIC) associated with malignancy has been recently suggested [35]. A recent study in an animal model further demonstrated that TFbearing MP derived from human cancer cells induced the activation of the coagulation cascade [34]. Moreover, MP derived from human pancreatic and lung cancer cells promoted thrombus formation after injection in an in vivo murine model, while endogenous cancer-derived MP contributed to the thrombogenic state in mice that developed ectopic tumors [45].

In summary, it is currently established that increased TF expression from cancer cells and the release of TF-bearing MP are implicated in the development of thromboembolic complications in patients with cancer.

#### 3.2.2 Myeloproliferative disorders (MPD)

MPDs are a group of clonal hematopoietic disorders, characterized by increased production of leukocytes, erythrocytes and/or platelets. Essential thrombocythemia (ET) and polycythemia vera (PV) are included in this group of syndromes. These disorders are frequently associated with arterial and venous thrombotic complications. The JAK2 (V617F) mutation, which is detectable in 90 - 95% of cases of PV and in only 50 - 60% of ET cases, has been correlated with an increased risk for thrombotic events in patients with PV and ET [46,47]. Moreover, increased TF expression in monocytes [48], platelets [49] and polymorphonuclear cells [50] from patients with MPD was associated with thrombotic complications. A correlation between TF expression and JAK2 (V617F) mutation was further suggested [48,49]. It is of interest that treatment with hydroxyurea reduced the levels of TF in neutrophils from patients with ET and PV, providing a further linkage between circulating TF expression and disease pathogenesis [50]. A recent study also demonstrated increased TF-bearing MP derived from platelets in patients with ET [38], even though a direct correlation between thrombotic events and levels of MP was not indicated.

#### 3.2.3 Antiphospholipid syndrome (APS)

Arterial and venous thrombotic events and recurrent miscarriages are common disease manifestations of APS. Even though the pathogenesis of this syndrome is not fully elucidated, accumulating evidence suggests a critical role for increased TF expression by blood cells in the prothrombotic tendency of APS [16,51]. It has been proposed that complement activation by antiphospholipid antibodies results in the generation of the anaphylatoxin C5a, which in turn induces TF production [16,51]. The enhanced circulating TF expression activates the coagulation cascade, resulting in thrombosis. On the other hand, TF expression in neutrophils has been associated with the obstetric manifestations of APS [52], as discussed in the following section.

#### 3.3 Sepsis

DIC is a detrimental complication of sepsis and is characterized by the formation of thrombi in the microvasculature, resulting in multi-organ failure. Induction of circulating TF expression, due to the proinflammatory microenvironment that characterizes sepsis, has been associated with DIC [53]. Several experimental models of endotoxemia provide evidence for this linkage. Increased levels of TF-bearing MP have been observed in a human experimental model of endotoxemia [54]. Similar results were extracted by an analogous murine model, suggesting the use of TF-bearing MP as a biomarker for the evaluation of the risk for DIC [55]. Recently, a study in a non-human primate model of Escherichia. coli-driven sepsis provided evidence for the implication of complement activation in this process. Complement inhibition reduced the activation of the coagulation cascade and the progression to DIC by decreasing the expression of TF [56].

#### 4. Non-thrombotic effect of TF-thrombin pathway

Several lines of evidence suggest a role for the TF-thrombin pathway in a growing spectrum of biological processes. It has been suggested that TF overexpression and the subsequent activation of serine proteases contribute to inflammation, tumor biology or fibrosis, implicating the coagulation cascade in the pathogenesis of various disorders. Signaling through PARs has been proposed as the mediating mechanism for the above mentioned effects. Four members (PAR1 - PAR4) constitute this family of G-protein-coupled receptors and are expressed in numerous cell types. The proteases of the extrinsic coagulation cascade can activate all four PARs. Thrombin is considered as a major activator of PAR1, PAR3 and PAR4, while TF-FVIIa targets PAR2 and FXa activates PAR1 and PAR2 [3]. Several intracellular pathways are activated downstream of PAR1 and PAR2 activation, including PI3K, Src family tyrosine kinases and the extracellular signalregulated kinase (ERK)/MAPK pathway for PAR1 and PI3K, ERK/MAPK and the G-proteins Rac and Rho-A for PAR2 [57]. PAR activation results in the expression and release of several cytokines and chemokines and is implicated in a variety of biological functions [3].

In the following section clinical observations and experimental data supporting the non-thrombotic functions of the TF-thrombin pathway are presented (Figures 1 and 2).

#### 4.1 Inflammatory responses

The TF-thrombin pathway has emerged as a significant player in inflammatory responses. The activation of this cascade seems to enhance inflammation in disease models including sepsis, ischemia-reperfusion (I/R) injury, arthritis and antiphospholipid syndrome (APS).

#### 4.1.1 Sepsis

The crosstalk between the coagulation cascade and inflammation in sepsis is bidirectional. The inflammatory milieu in sepsis is able to induce TF expression, resulting in the activation of coagulation cascade [53,54,56]. The subsequent fibrin deposition plays a key role in DIC, as described in the previous section [55,56]. On the other hand, the activation of TFthrombin pathway further induces inflammation, creating a vicious cycle.

Early studies in animal models demonstrated that inhibition of the TF-FVIIa complex by either active site-inhibited FVIIa (FVIIai), or tissue factor pathway inhibitor-1 (TFPI-1) resulted in decreased expression of the proinflammatory cytokines IL-6 and IL-8 and reduced mortality [57,58]. Activation of PARs was further implicated in the crosstalk between coagulation cascade and inflammatory responses. In an animal model for endotoxemia, both TF deficiency and combined inhibition of thrombin and deficiency in PAR2 reduced inflammation [59].

The beneficial effect of the TF-thrombin pathway inhibition in abolishing inflammation and preventing DIC in animal septic models led to the investigation of the effect of pharmaceutical intervention with the natural anticoagulants, TFPI-1, antithrombin-III (ATIII), and activated protein C (APC), in animal models of sepsis [57,58,60-62]. The promising results obtained from these studies prompted the initiation of clinical trials for the assessment of the effectiveness of these natural anticoagulants in sepsis, as discussed in a following section.

#### 4.1.2 Ischemia-reperfusion (I/R) injury

In addition to the key role of the coagulation cascade in atherothrombotic disease, the TF-thrombin pathway is involved in the inflammatory process associated with I/R, contributing in both morbidity and mortally of acute myocardial infarction and stroke. A study in an animal model revealed that the inhibition of TF and thrombin reduced the size of myocardial infarction in a manner independent of fibrin deposition. This study proposed that TF and thrombin mediate their effect through enhanced inflammation [63]. Moreover, fibrin-derived products have been implicated in I/R injury pathogenesis in a rat model of myocardial infarction [64]. Studies in murine models of myocardial I/R and cerebral ischemia further supported this suggestion, demonstrating the implication of PAR1 in myocardial remodeling and left ventricular hypertrophy [65] and in infract size, respectively [66]. Considering the effect of TF-FVIIa complex and PAR2 in this process, there are controversial reports. It has been recently shown that the inhibition of TF-FVIIa complex with FVIIai significantly reduced I/R injury in a murine model of myocardial I/R injury [67]. However, PAR2 activation has been shown to provide beneficial results in a rat model of myocardial I/R injury [68]. Despite the above experimental data, there no reported clinical data assessing the role of anticoagulation with the natural anticoagulants in I/R injury.

#### 4.1.3 Antiphospholipid syndrome (APS)

Recurrent miscarriages frequently complicate the disease course of APS. Pregnancy morbidity was originally thought to depend on thrombosis of the placenta. However, recent experimental data suggest a role for TF-FVIIa-complex-dependent induction of inflammation in this process.

Experimental evidence indicates that complement activation due to antiphospholipid antibodies results in the overexpression of TF in the surface of neutrophils [16,52]. TF-FVIIa signaling through PAR2 in neutrophils was demonstrated to induce oxidative stress. The resultant production of reactive oxygen species after PAR2 activation was further suggested to lead to trophoblast oxidative injury and pregnancy loss [69]. An interesting observation is that statins reduced both TF and PAR2 expression and prevented pregnancy loss [69]. However, it has to be demonstrated whether the use of statins is beneficial in clinical practice.

#### 4.1.4 Arthritis

Clinical and experimental evidence also implicates the TF-thrombin pathway in the pathogenesis of arthritis. Several studies demonstrate the activation of coagulation and fibrinolysis in synovial fluid from patients with rheumatoid arthritis [70]. Furthermore, increased functional TF activity was observed in synovial membranes from patients with rheumatoid arthritis compared with those from patients with osteoarthritis. This study further associated inhibition of TF signaling by FVIIai with decreased synovial inflammation as detected by the measurement of synovial thickness and articular cartilage damage in murine antigen-induced arthritis [71]. Signaling through PAR2 may be implicated in this process due to the observed inhibition of joint inflammation in PAR2-deficient mice in an adjuvant monoarthritis model of chronic inflammation [72].

#### 4.2 Tumor biology

Accumulating evidence correlates TF expression by malignant cells with many aspects of the pathogenesis of cancer. Increased TF expression has been associated with aggressive and metastatic disease in breast [44], colorectal [42] and pancreatic cancer [43,73]. Apart from the implication of TF in venous thrombosis in cancer patients, a key role for TF-FVIIa and thrombin in metastasis, angiogenesis and tumor growth has been suggested. Enhanced TF expression [41-44,73] and ectopic synthesis of FVII [74] by cancer cells has been reported and implicated in tumor progression.



The vast majority of experimental data in various models of malignancy indicates that TF-FVIIa-PAR2 signaling contributes in tumor neoangiogenesis mainly due to the induction of VEGF. TF expression was correlated with VEGF expression and microvessel density in resected pancreatic cancer [43], while silencing of TF gene retarded (although not abrogated) tumor growth in vivo due to inhibition of angiogenesis in colorectal cancer [75]. Moreover, TF inhibition and PAR2 deficiency reduced proliferation and migration of malignant glioma cells in vitro [76], while TF-FVIIa inhibition impaired hepatic metastasis in colorectal cancer models [77]. Finally, it has been demonstrated that the administration of an inhibitory antibody of direct TF-FVIIa signaling attenuated tumor growth in a murine model, while PAR2 deficiency resulted in similar inhibition of the TF-FVIIa complex [78]. asTF has also been reported to induce angiogenesis independent of PAR2 activation but dependent on integrin function [24].

While the TF-FVIIa complex contributes primarily to tumor growth and angiogenesis, thrombin generation seems to be implicated in the progression to metastatic disease. Thrombin generation results in the formation of fibrin and platelet-rich envelopes that protect metastatic cancer cells from clearance by natural killer cells [79]. Experimental data demonstrated that endogenous thrombin generation enhances the metastatic capacity of cancer cells through PAR1 signaling [80] or cathepsin D upregulation [81]. PAR1 silencing has been further reported to attenuate metastasis in malignant melanoma [82]. The implication of PAR1 in the metastatic process is also suggested by the increased expression of PAR1 in cancer cells [83] and in neoplastic cells from patients with metastatic melanoma cells compared with dysplastic nevi and primary melanoma [84].

#### 4.3 Fibrosis

The activation of the extrinsic coagulation cascade has been implicated in fibrosis as well. PAR-1 signaling has been shown to induce connective tissue growth factor (CTGF) expression. In accordance with this, the *in vitro* activation of lung fibroblasts from patients with systemic sclerosis for CTGF production and collagen deposition in a thrombin-dependent manner has been recently demonstrated. This effect was attenuated in fibroblasts treated with the thrombin inhibitor dabigatran [85].

Enhanced TF pathway activity has been reported in patients suffering from pulmonary fibrotic disorders [13,86]. The involvement of the coagulation pathway in pulmonary fibrosis is further indicated by the beneficial effect of anticoagulant therapy in patients with idiopathic pulmonary fibrosis [87].

Moreover, TF-FVIIa complex participates in wound healing, as reported by a recent study in a murine in vivo model [88]. In addition, impaired wound healing has been shown in a murine model of hemophilia B, which was partially restored after administration of FIX or FVIIa [89].

#### 5. Anticoagulation therapies

In this section, moving on from the well established antithrombotic effect of anticoagulation regimens, we will discuss the clinical benefit of such treatment in non-thrombotic manifestations of the previously mentioned disorders (Figure 3) (Table 1). On the other hand, treating physicians need to take into account the risk of bleeding with anticoagulation therapy, and administer anticoagulant agents when the benefits outweigh the risks.

#### 5.1 Natural anticoagulants

Beneficial effects have been documented for the natural anticoagulants TFPI, ATIII, and recombinant human activated protein C (APC) in animal models of sepsis [56,58,61] and in Phase II clinical trials [90-92]. However, these effects were confirmed only for human recombinant APC in a Phase III clinical trial [93]. This regimen is currently approved for the treatment of adult patients with severe sepsis with multiple organ failure in the USA and Europe. However, several concerns have been raised regarding the approval and the post-marketing surveillance of APC in the management of severe sepsis [94]. It thus remains to be further evaluated for which patient and at which specific time-point during the septic process APC is beneficial for the management of septic patients.

#### 5.2 TF-FVIIa inhibitors

Nematode anticoagulant protein c2 (NAPc2) is a potent anticoagulant that targets the TF-FVIIa complex, abrogating thrombin generation. It was originally isolated from a haematophagous hookworm, Ancylostoma caninum [95]. The safety and the effectiveness of the recombinant form of NAPc2, rNAPc2, in the treatment in patients undergoing elective coronary angioplasty and for the prevention of venous thromboembolism after total knee arthroplasty has been evaluated in Phase I clinical trials [96,97]. Moreover, rNAPc2 provided beneficial results in the treatment of patients with non-STsegment elevation acute coronary syndrome, while it was well tolerated [98].

Studies in experimental models evaluated the effect of the inhibition of TF-FVIIa complex signaling by rNAPc2 in cancer biology and inflammation. rNAPc2 has shown effectiveness in the inhibition of tumor growth and metastasis in an animal experimental model of colorectal cancer by inhibiting TF [99]. It is of interest that the administration of this agent in a primate model of Ebola hemorrhagic fever prolonged the survival of infected animals by attenuating the coagulation and proinflammatory response [100]. However, rNAPc2 failed to ameliorate the host defense in a murine model of pneumococcal pneumonia and a similar model of abdominal sepsis, despite the proper inhibition of TF-FVIIa activity [101,102].

TF-FVIIa complex is also inhibited by Ixolaris, a tick salivary protein with a function similar to TFPI. Experimental

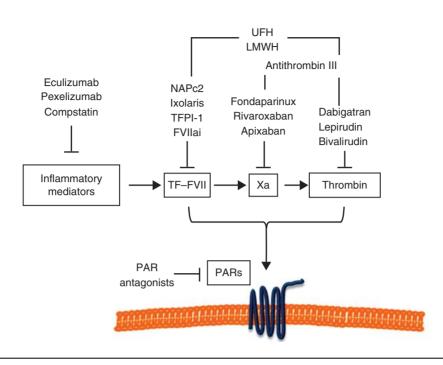


Figure 3. Targeting the TF-thrombin axis in non-thrombotic disorders. Several agents that interfere with the components of TF-thrombin axis have been developed. Clinical evidence suggests an antimetastatic effect of unfractionated heparin (UFH) and low-molecular-weight heparins (LMWH), while the data for their anti-inflammatory effect are inconclusive. Antithrombin III, TFPI and FVIIa inhibitors (FVIIai) have shown an anti-inflammatory potential in experimental models, which was not confirmed by clinical studies for the treatment of sepsis. Recombinant Nematode anticoagulant protein c2 (rNAPc2) and ixolaris are inhibitors of the TF-FVIIa complex, which have shown effectiveness as antimetastatic agents in experimental models. The anti-inflammatory and anti-metastatic effects of the recently developed factor Xa and thrombin inhibitors have not been extensively evaluated yet. The administration of agents that prevent the upregulation of TF expression by blocking complement activation or inhibit protease activated receptor (PAR) signaling may emerge as future strategies that target the TF-thrombin axis.

TFPI: TF pathway inhibitor.

data demonstrated a potent antithrombotic activity for ixolaris [103]. Moreover, a study in an experimental glioblastoma model provided evidence for the effectiveness of this agent in the inhibition of angiogenesis and tumor growth [104].

#### 5.3 Heparins

Heparins can be divided according to their structure and pharmacologic profiles into different groups: unfractionated heparin (UFH), low-molecular-weight heparins (LMWH) and the synthetic pentasaccharides fondaparinux and idraparinux [105,106].

Although UFH is highly efficacious in the treatment of venous thromboembolism (VTE), its clinical use has been restricted due to the intravenous route of administration, the need for continuous laboratory monitoring and the relatively common occurrence of heparin-induced thrombocytopenia (HIT) [105].

LMWH have replaced UFH in the treatment and prevention of venous and arterial thromboembolism. LMWH show similar efficacy to UFH, while they are administered subcutaneously in weight-adjusted fixed dosages with no need for laboratory monitoring, rendering feasible the administration

in the outpatient setting. However, the risk for HIT is still present with LMWH, even though in a significantly lower percentage of patients compared with UFH [105].

Fondaparinux and idraparinux are synthetic pentasaccharides that promote the inactivation of FXa by antithrombin. Fondaparinux is approved for the prevention and treatment of DVT and PE following orthopedic and abdominal surgery and for the treatment of acute coronary syndromes. Major bleeding events occurred infrequently, in a rate similar to that observed in patients under treatment with LMWH. Idraparinux has a very long half-time and it is administered subcutaneously once-weekly. Clinical trials have evaluated the efficacy of idraparinux in the treatment and secondary prophylaxis of DVT and PE and the long-term prevention of stroke in patients with atrial fibrillation [106].

Apart from their antithrombotic activity, an antiinflammatory role has been suggested for heparins. Experimental data demonstrated a beneficial effect of heparins in animal models of chronic inflammation, like colitis [107] or arthritis [108]. On the other hand, there are contradictory clinical data for the efficacy of UFH and LMWH in inflammatory disorders [109,110]. The anti-inflammatory effect



Table 1. Potential non anti-thrombotic effects of anti-coagulants.

	Targeted factors	Anti-metastatic effect	Anti-inflammatory effect
Heparins			
ÜFH	lla, Xa, IXa, Xla, Xlla	Experimental and clinical evidence [113-115]	Experimental and clinical evidence [108]
LMWH	lla, Xa	Experimental and clinical evidence [112-115]	Experimental evidence, contradictory clinical data [107,109-111]
Fondaparinux	Xa	No	No
Vitamin K inhibitors	II, VII, IX, X	No	No
New oral anticoagulants			
Dabigatran	lla	Not assessed	Not assessed (experimental evidence for anti-fibrotic action) [85]
Rivaroxaban	Xa	Not assessed	Not assessed
Apixaban	Xa	Not assessed	Not assessed
TF_FVIIa inhibitors			
NAPc2	TF_FVIIa	Experimental data [99]	Contradictory experimental data [101-102]
Ixolaris	TF-FVIIa	Experimental data [104]	No

LMWH: Low-molecular weight heparins: NAPc2: Nematode anticoagulant protein c2: UFH: Unfractionated heparin

of heparins is independent from their anti-coagulant activity and has been attributed to their ability to interact with selectins and to inhibit complement activation [111].

The interaction between the polysaccharide chain of heparins and selectins is suggested to mediate the anti-metastatic effect of heparins [112]. Treatment with LMWH has been shown to improve the survival in cancer patients [113]. Apart from the reduction of thrombosis-dependent morbidity and mortality, an independent-to-anticoagulation effect was suggested by the prolonged survival of patients with nonmetastatic cancer under treatment with LMWH compared with those receiving vitamin K antagonists. This effect was not observed in patients with metastatic cancer, suggesting an anti-metastatic effect for LMWH [113]. These clinical observations were confirmed by experimental studies in animal models [114,115]. The specificity of the interaction between heparins and selectins in their anti-metastatic potential is further indicated by the inability of fondaparinux, which lacks a polysaccharide chain, to attenuate metastasis in an experimental model [112].

#### 5.4 Vitamin K antagonists (VKA)

Warfarin and other VKA are effective in the primary and secondary prevention and treatment of VTE. However, the necessity for frequent laboratory monitoring, the drug interactions and the severe bleeding adverse events constitute limitations to their clinical use [116]. Moreover, an anti-metastatic effect was not demonstrated for VKA, as previously mentioned [113]. This observation, along with an increase in bleeding events in cancer patients receiving VKA and antineoplastic agents, render LMWH as a preferable treatment option in patients with malignancy [113].

It is of interest that a study in patients with idiopathic pulmonary fibrosis demonstrated a beneficial effect for

warfarin and LMWH in the progression of the disease [87]. The latter has to be further evaluated.

#### 5.5 New oral anticoagulants

The limitations in the clinical use of heparins and VKA prompted the development of new oral anticoagulants: dabigatran that specifically inhibits thrombin, and rivaroxaban and apixaban that inhibit FXa [106].

Phase III clinical trials have documented the efficacy of dabigatran in the prevention of VTE after orthopedic surgery [117] and in long-term treatment and secondary prevention of VTE [118]. Furthermore, in a Phase III clinical trial in patients with atrial fibrillation, dabigatran was associated with rates of stroke and systemic embolism that were similar to those associated with warfarin but lower rates of major hemorrhage at a dose of 110 mg, while at a dose of 150 mg was associated with lower rates of stroke and systemic embolism but similar rates of major hemorrhage as compared with warfarin [119]. Even though experimental data suggest a significant beneficial effect of thrombin inhibition in critical functions like inflammation and tumor metastasis, this effect has to be evaluated in clinical studies.

Rivaroxaban and apixaban are direct inhibitors of FXa given orally. These agents were superior to LMWH or warfarin in the prevention of VTE after orthopedic surgery [120,121]. Several data also demonstrate their effectiveness in the treatment of VTE [122,123]. Clinical trials are currently evaluating the effectiveness of these agents in the prevention of cardioembolic stroke in patients with atrial fibrillation [124].

#### 5.6 Hirudin analogues

Lepirudin and bivalirudin are included in this group of pharmacological agents. Both agents act as direct thrombin inhibitors.

Lepirudin is a recombinant analogue of the leech protein hirudin. Unlike heparins, it binds directly and irreversibly to thrombin, resulting in its neutralization. Lepirudin is used as an antithrombotic agent in patients with HIT [125].

Bivalirudin is a synthetic peptide which specifically binds and inhibits both circulating and clot-bound thrombin [126]. Clinical studies have demonstrated its effectiveness in the treatment of patients with acute coronary events undergoing percutaneous coronary intervention [127,128]. Moreover, the efficacy of bivalirudin was evaluated in patients with HIT during cardiopulmonary bypass [129].

#### 6. Conclusion

Several experimental and clinical lines of evidence currently suggest a critical role for the TF-VIIa complex in human biology. The description of TF-VIIa complex, FXa and thrombin signaling pathways implicates the coagulation cascade, apart from the well-established role in thrombosis, in the pathogenesis of inflammatory disorders and malignancy. This implication is the springboard for the investigation of the clinical effect of anticoagulation therapies in the treatment of the above mentioned disorders. Moreover, new anti-thrombotic regimens are nowadays available, which offer the opportunity for therapeutic intervention in both thrombotic and non-thrombotic coagulation-cascadedependent disorders. However, the risk of major bleeding events in patients under anticoagulation treatment is considerable and raises concerns for their use in the treatment for non thrombotic disorders. The development of agents with favorable safety profile and tolerability could enable the broader use of anticoagulants in the treatment of both thrombotic and non thrombotic disorders.

#### 7. Expert opinion

A new era for anti-coagulation therapeutic strategies emerges in view of the novel agents with improved pharmacokinetics and safety profiles. The efficacy of the recently developed regimens in the prevention and treatment of thromboembolism has been evaluated, providing alternative solutions. However, extensive basic and clinical research is needed to evaluate whether FXa and thrombin inhibitors exert possible antiinflammatory and/or anti-neoplastic effects as presumed from the currently available data that suggest a beneficial effect of thrombin and/or PAR inhibition or deficiency in experimental models of inflammation and cancer. Moreover, the development and application of PAR inhibitors in clinical practice may be an alternative prospective step in the treatment of diseases and syndromes like sepsis, I/R injury or cancer, which constitute a major challenge for the present therapeutic strategies.

Another option for intervening in the activity of the TF-thrombin pathway is the downregulation of the increased TF expression that is responsible for the thrombotic and

non-thrombotic manifestations observed in several disorders. The inhibition of the inflammatory mediators implicated in the induction of TF expression is a potential target for therapeutic intervention. In this direction, the inhibition of complement activation by the administration of the recently developed monoclonal antibodies targeting C5, pexelizumab and eculizumab, could be beneficial for a group of disorders whose pathogenesis depends on the cross-talk between C5a and TF. Patients under extra-corporal circulation, like patients undergoing hemodialysis or coronary artery bypass surgery, or patients with APS and sepsis frequently suffer from thrombotic manifestations associated, at least in part, with the increased TF expression driven by the generation of C5a. Recently, a study in a non-human primate model demonstrated that the inhibition of complement activation using compstatin prevented sepsis-induced coagulopathy by downregulating TF and attenuated multi-organ failure [56]. Moreover, a recent study reported that complement activation, induced by the biomaterials of hemodialysis, results in the overexpression of circulating TF, suggesting an implication in the pro-thrombotic state of end-stage renal disease patients. Ex vivo and in vitro studies suggested that treatment with compstatin precluded TF overexpression [17]. Complement inhibition with compstatin also prevented the induction of TF expression in leukocytes incubated with sera from patients with APS [16]. Considering APS, an animal model suggested a beneficial effect of treatment with statins in the obstetric manifestations of the syndrome via TF downregulation [52], suggesting an additional treatment strategy targeting TF. Moreover, the inhibition of C5a by pexelizumab ameliorated the prognosis in patients undergoing coronary artery bypass surgery, which could be attributed, at least in part, to the induction of TF expression [130]. In addition, the administration of eculizumab in patients with paroxysmal nocturnal hemoglobinuria had a beneficial effect on the thrombotic manifestations of the disease [131]. The imperative need for targeted therapeutic agents for the treatment of disorders with large effects on public health motivates the experimental efforts towards novel directions.

Another significant area of research is the identification of possible additional indications for anti-coagulation agents. Existing experimental and clinical data suggest a role for LMWH in the progression of cancer by attenuating metastasis. The elucidation of the anti-neoplastic mechanism of action of LMWH would break new ground in the investigation of the pathogenesis of metastasis and in the treatment of cancer. It would be of great interest to determine whether the administration of LMWH as an additional to chemotherapy strategy would ameliorate the disease course in all or a subgroup of patients with specific cell type or grade of malignancy, independently or not from the prevention of thrombotic events. The recently introduced anti-coagulation agents could contribute to the expansion of the therapeutic options in cancer patients, if proven effective, especially due to their improved safety profile.



In conclusion, further research is needed for the development of anti-coagulation agents with better safety profile and pleiotropic actions, which would reinforce our arsenal for the treatment of disorders with increasing morbidity and mortality.

#### **Declaration of interest**

The authors state no conflict of interest and have received no payment in preparation of this manuscript.

#### **Bibliography**

Papers of special note have been highlighted as either of interest (•) or of considerable interest ( o o ) to readers

- Bachli E. History of tissue factor. Br J Haematol 2000;110:248-55
- An interesting historical review that describes the key steps in the investigation of the TF pathway.
- Rapaport SI, Rao LV. The tissue factor pathway: how it has become a "prima ballerina". Thromb Haemost 1995;74:7-17
- Coughlin SR. Protease-activated receptors in hemostasis, thrombosis and vascular biology. J Thromb Haemost 2005:3:1800-14
- A review that describes the interplay between coagulation and protease-activated receptors.
- Bazan JF. Structural design and molecular evolution of a cytokine receptor superfamily. Proc Natl Acad Sci USA 1990;87:6934-6938
- Mackman N, Morrissey JH, Fowler B, Edgington TS. Complete sequence of the human tissue factor gene. a highly regulated cellular receptor that initiates the coagulation protease cascade. Biochemistry 1989;28:1755-62
- Morrissey JH, Fakhrai H, Edgington TS. Molecular cloning of the cDNA for tissue factor, the cellular receptor for the initiation of the coagulation protease cascade. Cell 1987;50:129-35
- Spicer EK, Horton R, Bloem L, et al. Isolation of cDNA clones coding for human tissue factor: primary structure of the protein and cDNA. Proc Natl Acad Sci USA 1987;84:5148-52
- 8. Brand K, Fowler BJ, Edgington TS, Mackman N. Tissue factor mRNA in THP-1 monocytic cells is regulated at both transcriptional and posttranscriptional levels in response to lipopolysaccharide. Mol Cell Biol 1991;11:4732-8
- Bogdanov VY, Balasubramanian V, Hathcock J, et al. Alternatively spliced human tissue factor: a circulating,

- soluble, thrombogenic protein. Nat Med 2003-9-458-62
- This study describes the identification and functional characterization of asTF.
- 10. Drake TA, Morrissey JH, Edgington TS. Selective cellular expression of tissue factor in human tissues. Implications for disorders of hemostasis and thrombosis. Am J Pathol 1989:134:1087-97
- Fleck RA, Rao LV, Rapaport SI, et al. Localization of human tissue factor antigen by immunostaining with monospecific, polyclonal anti-human tissue factor antibody. Thromb Res 1990:59:421-37
- 12. Wilcox JN, Smith KM, Schwartz SM, Gordon D. Localization of tissue factor in the normal vessel wall and in the atherosclerotic plaque. Proc Natl Acad Sci USA 1989:86:2839-43
- This paper reported the overexpression of TF in atherosclerotic plaques, initiating the investigation of its role in the pathogenesis of atherothrombosis.
- Kambas K, Markiewski MM, 13 Pneumatikos IA, et al. C5a and TNF-alpha up-regulate the expression of tissue factor in intra-alveolar neutrophils of patients with the acute respiratory distress syndrome. J Immunol 2008;180:7368-75
- 14. Rafail S, Ritis K, Schaefer K, et al. Leptin induces the expression of functional tissue factor in human neutrophils and peripheral blood mononuclear cells through JAK2-dependent mechanisms and TNFalpha involvement. Thromb Res 2008;122:366-75
- Parry GC, Mackman N. Transcriptional regulation of tissue factor expression in human endothelial cells. Arterioscler ThrombVasc Biol 1995:15:612-21
- 16. Ritis K, Doumas M, Mastellos D, et al. A novel C5a receptor-tissue factor cross-talk in neutrophils links innate

- immunity to coagulation pathways. J Immunol 2006;177:4794-802
- This study along with [51], [52] and [69] describe the crosstalk between C5a and TF expression in neutrophils in the pathogenesis of antiphospholipid syndrome.
- Kourtzelis I, Markiewski MM, Doumas M, et al. Complement anaphylatoxin C5a contributes to hemodialysis-associated thrombosis. Blood 2010;116:631-9
- Armesilla AL, Lorenzo E, Gomez del Arco P, et al. Vascular endothelial growth factor activates nuclear factor of activated T cells in human endothelial cells: a role for tissue factor gene expression. Mol Cell Biol 1999;19:2032-43
- Mackman N. Regulation of the tissue factor gene. Thromb Haemost 1997:78:747-54
- A review presenting the regulation of TF promoter.
- Rong Y, Hu F, Huang R, et al. Early growth response gene-1 regulates hypoxia-induced expression of tissue factor in glioblastoma multiforme through hypoxia-inducible factor-1-independent mechanisms. Cancer Res 2006;66:7067-74
- Moll T, Czyz M, Holzmuller H, et al. Regulation of the tissue factor promoter in endothelial cells. Binding of NF KB-, AP-1-, and Sp1-like transcription factors. J Biol Chem 1995;270:3849-57
- Mallat Z, Benamer H, Hugel B, et al. Elevated levels of shed membrane microparticles with procoagulant potential in the peripheral circulating blood of patients with acute coronary syndromes. Circulation 2000;101:841-3
- This study describes the implication of microparticles in the pathogenesis of acute coronary syndromes.
- Censarek P, Bobbe A, Grandoch M, et al. Alternatively spliced human tissue factor (asHTF) is not pro-coagulant. Thromb Haemost 2007;97:11-14
- van den Berg YW, van den Hengel LG, Myers HR, et al. Alternatively spliced



#### The multivalent activity of the tissue factor-thrombin pathway in thrombotic and non-thrombotic disorders

- tissue factor induces angiogenesis through integrin ligation. Proc Natl Acad Sci USA 2009;106:19497-502
- 25 Bach RR. Initiation of coagulation by tissue factor. CRC Crit Rev Biochem 1988-23-339-268
- 26. Mackman N, Tilley RE, Key NS. Role of the extrinsic pathway of blood coagulation in hemostasis and thrombosis. Arterioscler Thromb Vasc Biol 2007;27:1687-93
- A descriptive review of the role of TF in atherothrombosis and deep venous thrombosis.
- Monroe DM, Hoffman M, Roberts HR. Platelets and thrombin generation. Arterioscler Thromb Vasc Biol 2002;22:1381-9
- 28. Mackman N. The many faces of tissue factor. J Thromb Haemost 2009;7(Suppl 1):136-9
- Toschi V, Gallo R, Lettino M, et al. 29 Tissue factor modulates the thrombogenicity of human atherosclerotic plaques. Circulation 1997;95:594-9
- 30. Reininger AJ, Bernlochner I, Penz SM, et al. A 2-step mechanism of arterial thrombus formation induced by human atherosclerotic plaques. J Am Coll Cardiol 2010;55:1147-58
- An interesting study that presents the sequence of events for the formation of arterial thrombi and the involvement of TF in the second step of this sequence.
- Suefuji H, Ogawa H, Yasue H, et al. Increased plasma tissue factor levels in acute myocardial infarction. Am Heart J 1997;134:253-9
- 32. Day SM, Reeve JL, Pedersen B, et al. Macrovascular thrombosis is driven by tissue factor derived primarily from the blood vessel wall. Blood 2005;105:192-8
- Chou J, Mackman N, Merrill-Skoloff G, 33. et al. Hematopoietic cell-derived microparticle tissue factor contributes to fibrin formation during thrombus propagation. Blood 2004;104:3190-7
- Davila M, Amirkhosravi A, Coll E, et al. Tissue factor-bearing microparticles derived from tumor cells: impact on coagulation activation. J Thromb Haemost 2008;6:1517-724
- 35. Langer F, Spath B, Haubold K, et al. Tissue factor procoagulant activity of plasma microparticles in patients with

- cancer-associated disseminated intravascular coagulation. Ann Hematol 2008:87:451-7
- Hron G, Kollars M, Weber H, et al. Tissue factor-positive microparticles: cellular origin and association with coagulation activation in patients with colorectal cancer. Thromb Haemost 2007:97:119-23
- 37. Haubold K, Rink M, Spath B, et al. Tissue factor procoagulant activity of plasma microparticles is increased in patients with early-stage prostate cancer. Thromb Haemost 2009;101:1147-55
- Trappenburg MC, van Schilfgaarde M, Marchetti M, et al. Elevated procoagulant microparticles expressing endothelial and platelet markers in essential thrombocythemia. Haematologica 2009;94:911-18
- Sousou T, Khorana AA. New insights into cancer-associated thrombosis. Arterioscler Thromb Vasc Biol 2009:29:316-20
- Milsom C, Yu J, May L, et al. The role of tumor-and host-related tissue factor pools in oncogene-driven tumor progression. Thromb Res 2007;120(Suppl 2):S82-91
- Nakasaki T, Wada H, Shigemori C, et al. Expression of tissue factor and vascular endothelial growth factor is associated with angiogenesis in colorectal cancer. Am J Hematol 2002;69:247-54
- Khorana AA, Ahrendt SA, Ryan CK, et al. Tissue factor expression, angiogenesis, and thrombosis in pancreatic cancer. Clin Cancer Res 2007;13:2870-5
- Ueno T, Toi M, Koike M, et al. Tissue factor expression in breast cancer tissues: its correlation with prognosis and plasma concentration. Br J Cancer 2000;83:164-70
- Khorana AA, Francis CW, Menzies KE, et al. Plasma tissue factor may be predictive of venous thromboembolism in pancreatic cancer. J Thromb Haemost
- A paper that correlates TF with the prognosis of patients with cancer.
- Thomas GM, Panicot-Dubois L, Lacroix R, et al. Cancer cell-derived microparticles bearing P-selectin glycoprotein ligand 1 accelerate thrombus

- formation in vivo. J Exp Med 2009;206:1913-27
- 46. Vannucchi AM, Antonioli E, Guglielmelli P, et al. Prospective identification of high-risk polycythemia vera patients based on JAK2(V617F) allele burden. Leukemia 2007;21:1952-9
- Hsiao HH, Yang MY, Liu YC, et al. The association of JAK2V617F mutation and leukocytosis with thrombotic events in essential thrombocythemia. Exp Hematol 2007;35:1704-7
- Arellano-Rodrigo E, Alvarez-Larran A, Reverter JC, et al. Increased platelet and leukocyte activation as contributing mechanisms for thrombosis in essential thrombocythemia and correlation with the JAK2 mutational status. Haematologica 2006;91:169-75
- Arellano-Rodrigo E, Alvarez-Larran A, Reverter JC, et al. Platelet turnover, coagulation factors, and soluble markers of platelet and endothelial activation in essential thrombocythemia: relationship with thrombosis occurrence and JAK2 V617F allele burden. Am J Hematol 2009;84:102-8
- Maugeri N, Giordano G, Petrilli MP, et al. Inhibition of tissue factor expression by hydroxyurea in polymorphonuclear leukocytes from patients with myeloproliferative disorders: a new effect for an old drug? J Thromb Haemost 2006;4:2593-8
- Seshan SV, Franzke CW, Redecha P, et al. Role of tissue factor in a mouse model of thrombotic microangiopathy induced by antiphospholipid antibodies. Blood 2009;114:1675-83
- This study along with the [16] [52] and [69] describes the crosstalk between C5a and TF expression in neutrophils in the pathogenesis of antiphospholipid syndrome.
- Redecha P, Tilley R, Tencati M, et al. Tissue factor: a link between C5a and neutrophil activation in antiphospholipid antibody induced fetal injury. Blood 2007;110:2423-31
- This study along with [16], [51] and [69] describe the crosstalk between C5a and TF expression in neutrophils in the pathogenesis of antiphospholipid syndrome.
- Osterud B, Bjorklid E. The tissue factor pathway in disseminated intravascular coagulation. Semin Thromb Hemost 2001;27:605-17



- Aras O, Shet A, Bach RR, et al. 54. Induction of microparticle- and cell-associated intravascular tissue factor in human endotoxemia. Blood 2004;103:4545-53
- Wang JG, Manly D, Kirchhofer D, et al. Levels of microparticle tissue factor activity correlate with coagulation activation in endotoxemic mice. J Thromb Haemost 2009;7:1092-8
- Silasi-Mansat R, Zhu H, Popescu NI, 56 et al. Complement inhibition decreases the procoagulant response and confers organ protection in a baboon model of E. coli sepsis. Blood 2010;116:1002-10
- This study describes the beneficial effect of complement inhibition in progression towards DIC in septic non-human primates.
- Ramachandran R, Hollenberg MD. Proteinases and signalling: pathophysiological and therapeutic implications via PARs and more. Br J Pharmacol 2008;153(Suppl 1):S263-82
- Taylor F, Chang A, Peer G, et al. Active site inhibited factor VIIa (DEGR VIIa) attenuates the coagulant and interleukin-6 and -8, but not tumor necrosis factor, responses of the baboon to LD100 Escherichia coli. Blood 1998:91:1609-15
- Pawlinski R, Pedersen B, Schabbauer G, et al. Role of tissue factor and protease-activated receptors in a mouse model of endotoxemia. Blood 2004:103:1342-7
- Creasey AA, Chang AC, Feigen L, et al. Tissue factor pathway inhibitor reduces mortality from Escherichia coli septic shock, J Clin Invest 1993;91:2850-6
- Taylor FB Jr, Emerson TE Jr, Jordan R, et al. Antithrombin-III prevents the lethal effects of Escherichia coli infusion in baboons. Circ Shock 1998;26:227-35
- Taylor FB Jr, Chang A, Esmon CT, 62. et al. Protein C prevents the coagulopathic and lethal effects of Escherichia coli infusion in the baboon. J Clin Invest 1987;79:918-25
- Erlich IH, Boyle EM, Labriola I, et al. Inhibition of the tissue factor-thrombin pathway limits infarct size after myocardial ischemia-reperfusion injury by reducing inflammation. Am J Pathol 2000;157:1849-62

- Petzelbauer P. Zacharowski PA. 64 Miyazaki Y, et al. The fibrin-derived peptide Bbeta15-42 protects the myocardium against ischemia-reperfusion injury. Nat Med 2005;11:298-304
- Pawlinski R, Tencati M, Hampton CR, 65 et al. Protease-activated receptor-1 contributes to cardiac remodeling and hypertrophy. Circulation 2007;116:2298-306
- Junge CE, Sugawara T, Mannaioni G, 66 et al. The contribution of protease-activated receptor 1 to neuronal damage caused by transient focal cerebral ischemia. Proc Natl Acad Sci USA 2003-100-13019-24
- Loubele ST, Spek CA, Leenders P, et al. Active site inhibited factor VIIa attenuates myocardial ischemia/ reperfusion injury in mice. J Thromb Haemost 2009;7:290-8
- 68. Napoli C, Cicala C, Wallace JL, et al. Protease-activated receptor-2 modulates myocardial ischemia-reperfusion injury in the rat heart. Proc Natl Acad Sci USA 2000:97:3678-83
- Redecha P, Franzke CW, Ruf W, et al. Neutrophil activation by the tissue factor/Factor VIIa/PAR2 axis mediates fetal death in a mouse model of antiphospholipid syndrome. J Clin Invest 2008;118:3453-61
- This study along with the [16], [51] and [52] describe the crosstalk between C5a and TF expression in neutrophils in the pathogenesis of antiphospholipid syndrome.
- 70. So AK, Varisco PA, Kemkes-Matthes B, et al. Arthritis is linked to local and systemic activation of coagulation and fibrinolysis pathways. J Thromb Haemost 2003:1:2510-5
- Busso N, Morard C, Salvi R, et al. Role of the tissue factor pathway in synovial inflammation. Arthritis Rheum 2003;48:651-9
- Ferrell WR, Lockhart JC, Kelso EB, et al. Essential role for proteinase-activated receptor-2 in arthritis. J Clin Invest 2003;111:35-41
- Kakkar AK, Lemoine NR, Scully MF, et al. Tissue factor expression correlates with histological grade in human pancreatic cancer. Br J Surg 1995;82:1101-4
- Koizume S, Jin MS, Miyagi E, et al. Activation of cancer cell migration and

- invasion by ectopic synthesis of coagulation factor VII. Cancer Res 2006;66:9453-60
- Yu JL, May L, Lhotak V, et al. Oncogenic events regulate tissue factor expression in colorectal cancer cells: implications for tumor progression and angiogenesis. Blood 2005;105:1734-41
- This study describes the regulation of TF in cancer cells and its implication in disease progression.
- Gessler F, Voss V, Dutzmann S, 76 et al. Inhibition of tissue factor/ protease-activated receptor-2 signaling limits proliferation, migration and invasion of malignant glioma cells. Neuroscience 2010:165:1312-22
- Zerbib P, Grimonprez A, Corseaux D, et al. Inhibition of tissue factor-factor VIIa proteolytic activity blunts hepatic metastasis in colorectal cancer. J Surg Res 2009;153:239-45
- 78. Versteeg HH, Schaffner F, Kerver M, et al. Inhibition of tissue factor signaling suppresses tumor growth. Blood 2008:111:190-9
- Nieswandt B, Hafner M, Echtenacher B, Mannel DN. Lysis of tumor cells by natural killer cells in mice is impeded by platelets. Cancer Res 1999;59:1295-300
- 80. Nierodzik ML, Chen K, Takeshita K, et al. Protease-activated receptor 1 (PAR-1) is required and rate-limiting for thrombin-enhanced experimental pulmonary metastasis. Blood 1998;92:3694-700
- Hu L, Roth JM, Brooks P, et al. Thrombin up-regulates cathepsin D which enhances angiogenesis, growth, and metastasis. Cancer Res 2008;68:4666-73
- 82. Villares GJ, Zigler M, Wang H, et al. Targeting melanoma growth and metastasis with systemic delivery of liposome-incorporated protease-activated receptor-1 small interfering RNA. Cancer Res 2008;68:9078-86
- Even-Ram S, Uziely B, Cohen P, et al. Thrombin receptor overexpression in malignant and physiological invasion processes. Nat Med 1998;4:909-14
- Tellez CS, Davis DW, Prieto VG, et al. 84 Quantitative analysis of melanocytic tissue array reveals inverse correlation between activator protein-2alpha and protease-activated receptor-1 expression



#### The multivalent activity of the tissue factor-thrombin pathway in thrombotic and non-thrombotic disorders

- during melanoma progression. J Invest Dermatol 2007;127:387-93
- 85. Bogatkevich GS, Ludwicka-Bradley A, Silver RM. Dabigatran, a direct thrombin inhibitor, demonstrates antifibrotic effects on lung fibroblasts. Arthritis Rheum 2009;60:3455-64
- Gunther A, Mosavi P, Ruppert C, et al. 86. Enhanced tissue factor pathway activity and fibrin turnover in the alveolar compartment of patients with interstitial lung disease. Thromb Haemost 2000:83:853-60
- 87. Kubo H, Nakayama K, Yanai M, et al. Anticoagulant therapy and idiopathic pulmonary fibrosis. Chest 2005:128:1475-82
- Xu Z, Xu H, Ploplis VA, Castellino FJ. 88. Factor VII deficiency impairs cutaneous wound healing in mice. Mol Med 2010;16:167-76
- McDonald A, Hoffman M, Hedner U, et al. Restoring hemostatic thrombin generation at the time of cutaneous wounding does not normalize healing in hemophilia B. J Thromb Haemost 2007:5:1577-83
- 90. Abraham E, Reinhart K, Svoboda P, et al. Assessment of the safety of recombinant tissue factor pathway inhibitor in patients with severe sepsis: a multicenter, randomized, placebo-controlled, single blind, dose escalation study. Crit Care Med 2001;29:2081-9
- 91. Warren BL, Eid A, Singer P, et al. High-dose antithrombin III in severe sepsis: a randomized controlled trial. JAMA 2001;286:1869-78
- 92 Abraham E, Reinhart K, Opal S, et al. Efficacy and safety of tifacogin (recombinant tissue factor pathway inhibitor) in severe sepsis: a randomized controlled trial. JAMA 2003;290:238-47
- 93. Bernard GR, Vincent JL, Laterre PF, et al. Efficacy and safety of recombinant human activated protein C for severe sepsis. N Engl J Med 2001;344:699-709
- A phase III clinical study that introduced the clinical use of activated protein C in severe sepsis.
- 94 Poole D, Bertolini G, Garattini S. Errors in the approval process and post-marketing evaluation of drotrecogin alfa (activated) for the treatment of severe sepsis. Lancet Infect Dis 2009;9:67-72

- Lee AY, Vlasuk GP. Recombinant nematode anticoagulant protein c2 and other inhibitors targeting blood coagulation factor VIIa/tissue factor. J Intern Med 2003;254:313-21
- Moons AH, Peters RJ, Bijsterveld NR, et al. Recombinant nematode anticoagulant protein c2, an inhibitor of the tissue factor/factor VIIa complex, in patients undergoing elective coronary angioplasty. J Am Coll Cardiol 2003;41:2147-53
- Lee A, Agnelli G, Buller H, et al. Dose-response study of recombinant factor VIIa/tissue factor inhibitor recombinant nematode anticoagulant protein c2 in prevention of postoperative venous thromboembolism in patients undergoing total knee replacement. Circulation 2001;104:74-8
- Giugliano RP, Wiviott SD, Stone PH, et al. Recombinant nematode anticoagulant protein c2 in patients with non-ST-segment elevation acute coronary syndrome: the ANTHEM-TIMI-32 trial. J Am Coll Cardiol 2007;49:2398-407
- Zhao J, Aguilar G, Palencia S, et al. rNAPc2 inhibits colorectal cancer in mice through tissue factor. Clin Cancer Res 2009;15:208-16
- 100. Geisbert TW, Hensley LE, Jahrling PB, et al. Treatment of Ebola virus infection with a recombinant inhibitor of factor VIIa/tissue factor: a study in rhesus monkeys. Lancet 2003;362:1953-8
- 101. Rijneveld AW, Weijer S, Bresser P, et al. Local activation of the tissue factor-factor VIIa pathway in patients with pneumonia and the effect of inhibition of this pathway in murine pneumococcal pneumonia. Crit Care Med 2006:34:1725-30
- 102. Weijer S, Schoenmakers SH, Florquin S, et al. Inhibition of the tissue factor/factor VIIa pathway does not influence the inflammatory or antibacterial response to abdominal sepsis induced by Escherichia coli in mice. J Infect Dis 2004-189-2308-17
- 103. Nazareth RA, Tomaz LS, Ortiz-Costa S, et al. Antithrombotic properties of Ixolaris, a potent inhibitor of the extrinsic pathway of the coagulation cascade. Thromb Haemost 2006;96:7-13
- 104. Carneiro-Lobo TC, Konig S, Machado DE, et al. Ixolaris, a tissue factor inhibitor, blocks primary tumor growth and angiogenesis in a

- glioblastoma model. J Thromb Haemost 2009;7:1855-64
- 105. Segal JB, Streiff MB, Hofmann LV, et al. Management of venous thromboembolism: a systematic review for a practice guideline. Ann Intern Med 2007:146:211-22
- 106. Franchini M, Mannucci PM. A new era for anticoagulants. Eur J Intern Med 2009;20:562-8
- A recent review that presents the recently developed anticoagulants.
- 107. Wan MX, Liu Q, Wang Y, Thorlacius H. Protective effect of low molecular weight heparin on experimental colitis: role of neutrophil recruitment and TNF-alpha production. Inflamm Res 2002;51:182-7
- Lider O, Baharav E, Mekori YA, et al. Suppression of experimental autoimmune diseases and prolongation of allograft survival by treatment of animals with low doses of heparins. J Clin Invest 1989;83:752-6
- de Bievre MA, Vrij AA, Schoon EJ, et al. Randomized, placebo controlled trial of low molecular weight heparin in active ulcerative colitis. Inflamm Bowel Dis 2007-13-753-8
- 110. Torkvist L, Thorlacius H, Sjoqvist U, et al. Low molecular weight heparin as adjuvant therapy in active ulcerative colitis. Aliment Pharmacol Ther 1999;13:1323-38
- 111. Rao NV, Argyle B, Xu X, et al. Low anticoagulant heparin targets multiple sites of inflammation, suppresses heparin-induced thrombocytopenia, and inhibits interaction of RAGE with its ligands. Am J Physiol Cell Physiol 2010;299:C97-110
- Stevenson JL, Choi SH, Varki A. Differential metastasis inhibition by clinically relevant levels of heparins-correlation with selectin inhibition, not antithrombotic activity. Clin Cancer Res 2005;11:7003-11
- This paper proposes that the antimetastatic effect of LMWH is associated with selectin inhibition and is independent from their antithrombotic activity.
- 113. Kuderer NM, Khorana AA, Lyman GH, Francis CW. A meta-analysis and systematic review of the efficacy and safety of anticoagulants as cancer treatment: impact on survival and



- bleeding complications. Cancer 2007;110:1149-61
- An interesting meta-analysis of the effectiveness of heparins and vitamin K antagonists with the outcome of patients with cancer, suggesting the beneficial effect of LMWH.
- 114. Niers TM, Klerk CP, Di Nisio M, et al. Mechanisms of heparin induced anti-cancer activity in experimental cancer models. Crit Rev Oncol Hematol 2007;61:195-207
- 115. Smorenburg SM, Van Noorden CJ. The complex effects of heparins on cancer progression and metastasis in experimental studies. Pharmacol Rev 2001;53:93-105
- 116. Ansell J, Hirsh J, Poller L, et al. The pharmacology and management of the vitamin K antagonists: the Seventh ACCP Conference on Antithrombotic and Thrombolytic Therapy. Chest 2004;126(3 Suppl):204S-33S
- 117. Eriksson BI, Dahl OE, Rosencher N, et al. Dabigatran etexilate versus enoxaparin for prevention of venous thromboembolism after total hip replacement: a randomised, double-blind, noninferiority trial. Lancet 2007;370:949-56
- 118. Schulman S, Kearon C, Kakkar AK, et al. Dabigatran versus warfarin in the treatment of acute venous thromboembolism. N Engl J Med 2009;361:2342-52
- 119. Connolly SJ, Ezekowitz MD, Yusuf S, et al. Dabigatran versus warfarin in patients with atrial fibrillation. N Engl J Med 2009;361:1139-51
- 120. Lassen MR, Raskob GE, Gallus A, et al. Apixaban versus enoxaparin for thromboprophylaxis after knee replacement (ADVANCE-2): a randomised double-blind trial. Lancet 2010;375:779-80

- 121. Lassen MR, Ageno W, Borris LC, et al. Rivaroxaban versus enoxaparin for thromboprophylaxis after total knee arthroplasty. N Engl J Med 2008;358:2776-86
- 122. Buller HR, Agnelli G, Cohen A, et al. A dose-ranging study evaluating once-daily oral administration of the factor Xa inhibitor rivaroxaban in the treatment of patients with acute symptomatic deep vein thrombosis: the Einstein-DVT Dose- Ranging Study. Blood 2008;112:2242-7
- 123. Buller H, Deitchman D, Prins M, et al. Efficacy and safety of the oral direct factor Xa inhibitor apixaban for symptomatic deep vein thrombosis. The Botticelli DVT dose-ranging study. I Thromb Haemost 2008;6:1313-18
- 124. ROCKET AF Study Investigators. Rivaroxaban-once daily, oral, direct factor Xa inhibition compared with vitamin K antagonism for prevention of stroke and Embolism Trial in Atrial Fibrillation: rationale and design of the ROCKET AF study. Am Heart J 2010;159:340-347.e1
- 125. Lubenow N, Eichler P, Lietz T, Greinacher A. Lepirudin in patients with heparin-induced thrombocytopenia: results of the third prospective study (HAT-3) and a combined analysis of HAT-1, HAT-2, and HAT-3. J Thromb Haemost 2005;3:2428-36
- 126. Sciulli TM, Mauro VF. Pharmacology and clinical use of bivalirudin Ann Pharmacother 2002;36:1028-41
- 127. Parodi G, Migliorini A, Valenti R, et al. Comparison of bivalirudin and unfractionated heparin plus protamine in patients with coronary heart disease undergoing percutaneous coronary intervention (from the Antithrombotic Regimens aNd Outcome [ARNO] trial). Am J Cardiol 2010;105:1053-9

- 128. Mehran R, Lansky AJ, Witzenbichler B, et al. Bivalirudin in patients undergoing primary angioplasty for acute myocardial infarction (HORIZONS-AMI): 1-year results of a randomised controlled trial. Lancet 2009:374:1149-59
- Koster A, Dyke C, Aldea G, et al. Bivalirudin during cardiopulmonary bypass in patients with previous or acute heparin-induced thrombocytopenia and heparin antibodies: results of the CHOOSE-ON Trial. Ann Thorac Surg 2007:83:572-7
- Testa L, Van Gaal WJ, Bhindi R, et al. Pexelizumab in ischemic heart disease: a systematic review and meta-analysis on 15,196 patients. J Thorac Cardiovasc Surg 2008;136:884-93
- 131. Hillmen P, Muus P, Duhrsen U, et al. Effect of the complement inhibitor eculizumab on thromboembolism in patients with paroxysmal nocturnal hemoglobinuria. Blood 2007;110:4123-8

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